

## Oligomeric Flavanoids. Part 25<sup>a</sup>. Cleavage of the Acetal Functionality in A-type Proanthocyanidins

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**Abstract.** The hepta-*O*-methyl ethers **3** and **4** of procyanidin A-1 **1** and A-2 **2** are subject to facile cleavage of the acetal functionality with sodium cyanoborane in trifluoroacetic acid at 0°C. This straight forward chemical method permits the unambiguous establishment of the absolute configuration of the DEF-flavanyl unit and the D-ring carbon and oxygen atoms that are involved in the double linkage of the A-class proanthocyanidins.

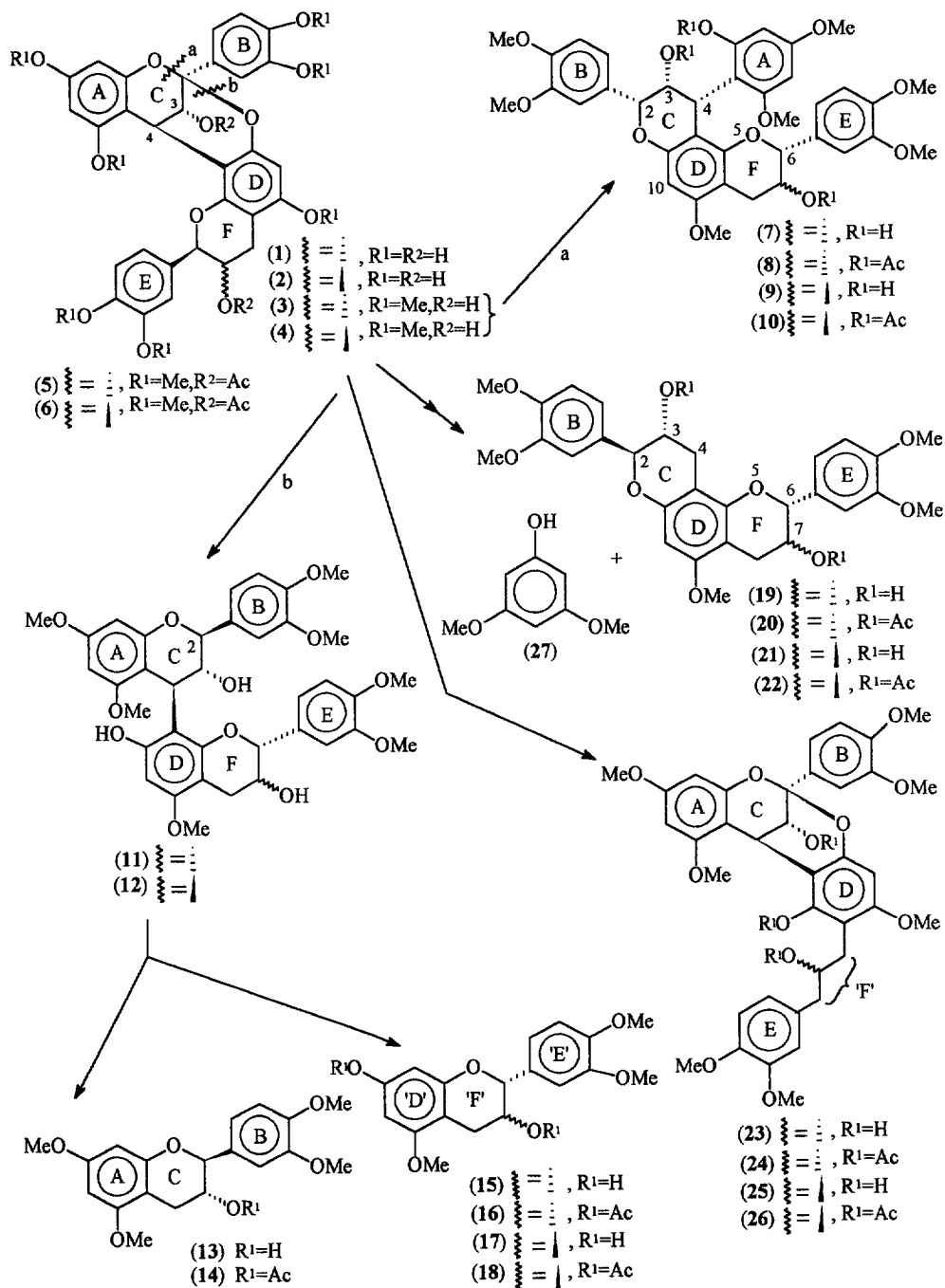
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The double interflavanyl linkage in A-type proanthocyanidins introduces a high degree of conformational stability which culminates in high-quality and unequivocal NMR spectra, conspicuously free of the effects of dynamic rotational isomerism at the dimeric level. Compounds of this class are readily recognizable from the characteristic AB-doublet ( $^3J_{3,4} = 3-4$  Hz) of C-ring protons in the heterocyclic region of their  $^1\text{H}$  NMR spectra<sup>1</sup>, and may possess either (2 $\alpha$ ,4 $\alpha$ )- or (2 $\beta$ ,4 $\beta$ )-double interflavanyl bonds. Two fundamental structural problems, *i.e.* establishment of the mode of linkage of the D- to the C-ring, and assignment of the absolute configuration at the stereocentres of the F-ring, have however limited progress in this field. These and related problems have hitherto been approached *via* exotic spectroscopic methods<sup>2-5</sup> which prompted us to search for a more simple and general chemical method that is based upon the reductive cleavage of the acetal functionality. The potential to address the aforementioned problems by reduction of either of the C-O acetal bonds is now demonstrated for the known procyanidins A-1 **1** and A-2 **2**, available from the skins of mature peanuts (*Arachis hypogaea* L.)<sup>6</sup>, using sodium cyanoborane [ $\text{Na}(\text{CN})\text{BH}_3$ ] in trifluoroacetic acid (TFA)<sup>7</sup>. The readily accessible hepta-*O*-methyl ethers **3** and **4** were selected as model compounds with a view to using the *O*-substituents of the D-ring as probes for  $^1\text{H}$  NMR studies.

### RESULTS AND DISCUSSION

Separate treatment of the hepta-*O*-methylprocyanidins A-1 **3** and A-2 **4** with  $\text{Na}(\text{CN})\text{BH}_3$  (1:2.4 molar ratio) in TFA for 1.5h at 0°C under  $\text{N}_2$  (Scheme), gave conversion to mixtures comprising the starting materials (**3**, 3.4% and **4**, 4.4% respectively), and as anticipated from cleavage 'a' the tetrahydropyrano[2,3-*f*]chromene derivatives (**9**, 5.2% and **7**, 7% respectively). The envisaged biflavanoids **11** and **12** from the 'b' pathway were,

<sup>a</sup> Part 24. Saunders, C.M.; Bonnet, S.L.; Steynberg, J.P.; Ferreira, D. *Tetrahedron*, 1996, **52**, 6003



**Scheme.** Cleavage of the acetal functionality of procyanidin A-1 and A-2 hepta- *O*-methyl ethers 3 and 4 with  $\text{Na}(\text{CN})\text{BH}_3$  in THF

however, not obtained but instead, the respective monomeric units, *i.e.* tetra-*O*-methyl-*ent*-catechin (**13**, 4%) and tri-*O*-methylcatechin (**17**, 3.4%) from the A-1 derivative **3**, and tetra-*O*-methyl-*ent*-catechin (**13**, 3%) and tri-*O*-methylepicatechin (**15**, 1.3%) from the A-2 derivative **4** were isolated (*vide infra*). These compounds were accompanied by the tetrahydropyrano[2,3-*f*]chromene derivatives (**21**, 3.9% and **19**, 4.5% respectively), the doubly linked epicatechin-1,3-diarylpropan-2-ol derivatives (**25**, 2.6% and **23**, 2.5% respectively), and 3,5-dimethoxyphenol (**27**, 3.8 and 4% respectively). The formation of compounds **19**, **21**, **23**, **25**, and **27** became more prominent under more drastic conditions.

The structures of the aforementioned products were unequivocally elucidated by comparison of the physical data of their *O*-acetyl derivatives, *e.g.* **8**, with those of similar derivatives of suitable reference compounds from our collection of related substances<sup>8-10</sup>. <sup>1</sup>H NMR data are collated in Tables 1 and 2 while circular dichroic (CD) data are given in the Experimental.

Owing to their importance to the protocol developed here, the structures of the tetrahydropyrano[2,3-*f*]chromene derivatives **8** and **10**, and of the flavan-3-ol derivatives **14**, **16** and **18** should be emphasized. Whereas the high amplitude negative Cotton effect ( $[\theta]_{239.5} -75000$ ;  $[\theta]_{242.4} -110000$  for **8** and **10** respectively) in the CD spectra of the tetrahydropyrano[2,3-*f*]chromene derivatives **8** and **10** are reminiscent of the 4 $\alpha$  orientation of their A-rings<sup>8</sup>, the 2,3-*cis*-3,4-*cis* relative configurations were evident from the <sup>1</sup>H NMR coupling constants of C-ring protons<sup>9</sup> (<sup>3</sup>J<sub>2,3</sub> = *ca* 1.0; <sup>3</sup>J<sub>3,4</sub> = 6.5 Hz for both **8** and **10**). The 2,4-*cis* relationship of A- and B-rings in these compounds was unequivocally established by the observed NOE associations<sup>11</sup> between 2-H(C) and 3-H(C) (7.3 and 7.0% respectively), and of 2-H(C) with 4-H(C) (5.4% for both **8** and **10**). In compound **10** the 2,4-*cis* orientation of A- and B-rings was additionally supported by the NOE effects between 2-OAc(A) and both 2-H(B) (0.9%) and 6-H(B) (0.7%). Although similar NOE's were absent in compound **8**, this derivative displayed an NOE-interaction between 2-OAc(A) and both 2- and 6-H(E) (1.1 and 1.0% respectively), thus confirming the 4,6-*cis* orientation of the A- and E-rings. Coupling constants of the F-ring protons (<sup>3</sup>J<sub>6,7</sub> = *ca* 1.0 and 9.0 Hz for **8** and **10** respectively) were indicative of a 6,7-*cis* and 6,7-*trans* configuration for compounds **8** and **10** respectively. Collectively these data confirmed the absolute configuration at the stereocentres as indicated in the tetrahydropyrano[2,3-*f*]chromene derivatives **8** and **10**, and especially the *R* configuration at C-3. The 5-*O*-methyl-7-*O*-acetyl substitution pattern of the A-ring of the epicatechin and catechin derivatives **16** and **18** was established by the observed NOE associations of 6-H(A) ( $\delta$  6.21 for both **16** and **18**) with both 5-OMe ( $\delta$  3.78, 3.76 for **16** and **18** respectively) and 7-OAc ( $\delta$  1.90, 1.95 for **16** and **18** respectively). Comparison of CD data of the flavan-3-ol derivatives **14**, **16** and **18** with those of the 3',4',5,7-tetra-*O*-methyl-3-*O*-acetyl derivatives of *ent*-catechin, epicatechin and catechin respectively, confirmed the 2*S*,3*R* absolute configuration of compound **13**, 2*R*, 3*R* of **15** and 2*R*,3*S* of **17**. These three flavan-3-ols originated from the reductive cleavage of the interflavanyl bond in the intermediate B-type procyanidin derivatives **11** and **12** (see also below).

**Table 1.**  $^1\text{H}$  NMR peaks ( $\delta_{\text{H}}$ ) of the tetrahydropyranochromene derivatives **8**, **10**, **20** and **22** and the doubly linked epicatechin derivatives **24** and **26** at 300 MHz (23°C) in  $\text{CDCl}_3$ . Splitting patterns and J-values(Hz) are given in parentheses.

Proton	<b>8</b>	<b>10</b>	<b>20</b>	<b>22</b> ( $\text{C}_6\text{D}_6$ )	<b>24</b>	<b>26</b>
3/6-H(A) 5/8-H(A)	6.37(d,2.5) 6.27(d,2.5)	6.15(d,2.5) 6.08(d,2.5)	-	-	6.31,6.10(each d,2.5)	6.28,5.94(each d,2.5)
2-H(B) 5-H(B) 6-H(B)	7.01(d,2.0) 6.82(d,8.5) 6.98(dd,2.0,8.5)	7.00(d,2.0) 6.80(d,8.5) 6.96(dd,2.0,8.5)	} 7.04-6.79	7.12(d,2.0) 6.63(d,8.5) 7.16(dd,2.0,8.5)	} 7.23-6.59	} 7.30-6.52
2-H(C) 3-H(C) 4-H(C)	5.02(br.s) 5.77(dd,1.0,6.5) 5.07(d,6.5)	5.01(br.s) 5.60(dd,1.0,6.5) 5.01(d,6.5)	4.98(d,7.0) 5.33(m) 3.10-2.71 (2xdd,1xm)	5.40(d,6.5) 5.77(m) 3.39(dd,6.0, 10.5) 3.15(dd,6.0,10.5)	- 5.48(d,3.0) 4.30(broadened)	- 5.57(d,3.0) 4.89(d,3.0)
6/10-H(D)	6.26(s)	6.25(s)	6.21(s)	6.48(s)	6.48(s)	6.18(s)
2-H(E) 5-H(E) 6-H(E)	6.47(d,2.0) 6.54(d,8.5) 6.03(dd,2.0, 8.5)	6.55(d,2.5) 6.58(d,8.5) 6.31(dd,2.5, 8.5)	} 7.04-6.79	7.03(d,2.2) 6.64(d,8.5) 7.05(dd,2.2, 8.5)	} 7.23-6.59	} 7.30-6.52
6-H(F)/CH <sub>2</sub> 7-H(F)	4.80(br.s) 5.23(m)	4.51(d,9.0) 5.02(m)	5.00(br.s) 5.48(br.s)	5.24(d,6.5) 5.77(m)	5.20(m)	5.34(m)
8 $\alpha$ -H(F) 8 $\beta$ -H(F)	} 2.90(d,3.5)	3.17(dd,6.0,16.0) 2.50(dd,9.5,16.0)	} 3.10-2.71 (2xdd,1xm)	3.34(dd,6.0,10.5) 3.10(dd,6.0,10.5)	} 3.00-2.42 (2xCH <sub>2</sub> )	} 2.78-2.15 (2xCH <sub>2</sub> )
OMe	3.89(3-B),3.85 (4-B),3.82(4-E), 3.78(9-D),3.76 (4-A),3.72(3-E), 3.62(6-A), each s	3.88(3-B),3.84 (4-B),3.82(4-E), 3.77(9-D),3.73 (4-A,3-E),3.46 (6-A), each s	3.89-3.78(x5), each s	3.52(3-E),3.50 (3-B),3.45(4-B, 4-E),3.40(10-D), each s	3.91-3.70(x7) each s	3.91,3.89,3.81, 3.75,3.69,3.41, 3.19,each s
OAc	1.62,1.70,1.82 (2-A),each s	1.64,1.71,1.79 (2-A),each s	1.93,1.92,each s	1.67,1.58,each s	2.39(D, br.s), 1.82(br.s) 1.73(s)	2.09(D),1.71, 1.55,each s

**Table 2.**  $^1\text{H}$  NMR peaks ( $\delta_{\text{H}}$ ) of the flavan-3-ol derivatives **14**, **16** and **18** at 300 MHz (23°C) in  $\text{CDCl}_3$ . Splitting patterns and J-values (Hz) are given in parentheses

Proton	<b>14</b>	<b>16</b>	<b>18</b>
6-H(A) 8-H(A)	6.07(d,2.5) 6.15(d,2.5)	6.21(d,2.0) 6.39(d,2.0)	6.21(d,2.0) 6.35(d,2.0)
2-H(B) 5-H(B) 6-H(B)	6.86(d,2.0) 6.80(d,8.0) 6.90(dd,2.0,8.0)	7.00(d,2.0) 6.84(d,8.5) 6.93(dd,2.0,8.5)	6.85(d,2.0) 6.80(d,8.0) 6.88(dd,2.0,8.0)
2-H(C) 3-H(C) 4 $\alpha$ -H(C) 4 $\beta$ -H(C)	5.0(d,7.0) 5.33(m) 2.89(dd,5.5,16.5) 2.65(dd,6.5,16.5)	5.0(br.s) 5.42(m) } 2.94(d,3.0)	5.02(d,7.0) 5.33(m) 2.90(dd,5.5,17.0) 2.67(dd,6.5,17.0)
OMe	3.85(4-B),3.84(3-B),3.76(5-A), 3.75(7-A), each s	3.88(4-B),3.87(3-B),3.78(5-A), each s	3.85(4-B),3.83(3-B),3.76(5-A), each s
OAc	1.94(3-C)(s)	1.90(3-C),2.28(7-A),each s	1.95(3-C),2.27(7-A),each s

The  $^1\text{H}$  NMR spectra (Table 1) of the tetrahydropyrano[2,3-*f*]chromene derivatives **20** and **22** each exhibited two ABM spin systems and a singlet in the aromatic region, two heterocyclic ABXY systems, five *O*-methyl and two *O*-acetyl resonances. Both compounds thus lack the oxygenated phenyl substituent at C-4 when compared to the 'normal' tetrahydropyrano[2,3-*f*]chromene derivatives **8** and **10**. Owing to the overlapping of some key *O*-methyl resonances, the two very similar aromatic ABM systems could not be differentiated in derivative **20**<sup>12</sup>. In the  $^1\text{H}$  NMR spectra (Table 1) of the doubly linked epicatechin-1,3-diarylpropan-2-ol derivatives **24** and **26** partial rotational barriers were observed at 23<sup>o</sup>C, but at 80<sup>o</sup>C nine aromatic and seven heterocyclic protons as well as seven *O*-methyl and three *O*-acetyl resonances were evident. The characteristic signals of the intact doubly linked epicatechin moiety (AB-doublet,  $^3J_{3,4} = 3.5$  Hz for both **24** and **26**) in conjunction with the one-proton multiplet and two sets of benzylic methylene protons confirmed the proposed structures. Since compounds **20**, **22**, **24** and **26** do not contribute significantly towards the methodology developed here, their structure elucidation need not be discussed further.

Both the carbon-oxygen bonds of the acetal functionality in the procyanidin A-1 and A-2 derivatives **3** and **4** are thus susceptible to reductive cleavage under acidic conditions. This process is presumably triggered by the random protonation of the acetal oxygens and concomitant delivery of the equivalent of a hydride ion at the antibonding ( $\sigma^*$ ) orbitals of the carbon-oxygen bonds in a predominant S<sub>N</sub>2 manner. Such a transfer of hydride ion apparently occurs from a complex between the reducing agent and the axial C-3 (C-ring) oxygen lone pair, the proximity of the boron-hydrogen bonds to the backside of the acetal carbon atom being a prerequisite for reduction of either one of the acetal bonds. Reduction thus leads to 'inversion' of configuration at C-2(C) of both B-type procyanidin intermediates **11** and **12**, and of the tetrahydropyrano[2,3-*f*]chromene derivatives **7** and **9**.

Biflavanoids **11** and **12** are prone to facile cleavage of their interflavanyl bonds *via* protonation of the electron-rich phloroglucinol D-ring<sup>14</sup> and attack of hydride ion at C-4(C)<sup>15</sup> to give the *ent*-catechin derivative **13** from the ABC-unit and respectively the epicatechin and catechin derivatives **15** and **17** from the DEF-moieties<sup>13</sup>. The doubly linked epicatechin-1,3-diarylpropan-2-ols **23** and **25** presumably results from the reductive cleavage of the benzyl ether functionality (F-ring) of hepta-*O*-methylprocyanidins A-1 **3** and A-2 **4**, in a process similar to the formation of 1,3-diarylpropanes when flavanones are treated with Na(CN)BH<sub>3</sub> in TFA<sup>16</sup>. The functionalized tetrahydropyrano[2,3-*f*]chromenes **7** and **9** may serve as the precursors to the tetrahydropyrano[2,3-*f*]chromene derivatives **19** and **21** *via* acid-catalyzed reductive rupturing of the labile C-10—phloroglucinol A-ring bond, thus also explaining the formation of 3,5-dimethoxyphenol **27**. The observed inversion of configuration at C-2(C) in compounds **19** and **21**, compared to that of the same stereocentres in the tetrahydropyrano[2,3-*f*]chromenes **7** and **9**, may indicate that the transfer of hydride ion in the acetal cleavage partially followed an S<sub>N</sub>1 pathway. It may also be effected *via* acid-catalyzed epimerization to afford the thermodynamically more stable 2,3-*trans* isomer. The latter process presumably occurs at a late stage when the diminished concentration of

$\text{Na}(\text{CN})\text{BH}_3$  does not permit hydride transfer to the equivalent of an incipient C-2-carbocation which is a prerequisite for epimerization.

The 'liberation' of the chain terminating flavan-3-ol units **15** and **17** unambiguously defines the D-ring oxygen that is involved in the acetal functionality of the parent compounds **1** and **2**. It furthermore provides a powerful probe towards addressing the hitherto unsolved problem of establishing the absolute configuration at the stereocentres of this moiety in naturally occurring A-type proanthocyanidins. The flavan-3-ol unit **13**, albeit with inversed C-2 configuration should facilitate the assignment of the absolute configuration at C-3 of the parent compounds **1** and **2**, especially in view of the inability to differentiate between 3,4-*cis*- and 3,4-*trans*-configuration in these compounds on the basis of  $^3J_{\text{HH}}$  values<sup>2</sup>. The mode of the C-C linkage between the constituent flavan-3-ol units in the A-type procyanidin, *e.g.* (4,6) or (4,8), is defined by the nature of the tetrahydropyranochromene, *i.e.* [2,3-*f*], [2,3-*g*] or [2,3-*h*], that is formed *via* reductive cleavage 'a'. The tetrahydropyranochromenes of type **8** represent a class of compounds with a well established method of structure elucidation<sup>8,12,17</sup>. This development should thus contribute substantially towards a straight forward chemically orientated structure definition of proanthocyanidins of the A-class.

## EXPERIMENTAL

<sup>1</sup>H NMR spectra were recorded on a Bruker AM-300 spectrometer for solutions in  $\text{CDCl}_3$  with  $\text{Me}_4\text{Si}$  as internal standard. J-values are given in Hz. Accurate mass estimations were obtained with a Varian CH-5 instrument with double focus. CD data were obtained in methanol on a JASCO J-710 spectropolarimeter. Preparative layer chromatography (PLC) was performed on plates (20x20 cm) with Merck Kieselgel PF<sub>254</sub> (1.0 mm) which were air-dried and used without prior activation. Acetylations were done in acetic anhydride/pyridine at ambient temperature. Evaporations were done under reduced pressure at *ca* 50°C in a rotary evaporator.

*Reductive cleavage of procyanidin A-2 hepta-O-methyl ether 4.* —  $\text{Na}(\text{CN})\text{BH}_3$  (37 mg,  $5.0 \times 10^{-4}$  mole) was added in portions over 30 min to a solution of the title compound **4** (50 mg,  $7.57 \times 10^{-5}$  mole) in TFA (3  $\text{cm}^3$ ) at 0°C under  $\text{N}_2$ . After 1h the reaction was quenched by the careful addition of water (20  $\text{cm}^3$ ) and the pH was adjusted to *ca* 6.9 (Merck special indicator, pH 4.0-7.0) with *aq.*  $\text{NaHCO}_3$  (2%). The mixture was extracted with ethyl acetate (3x50  $\text{cm}^3$ ) and the combined extract was stirred for 15 min with 3 drops of a solution of tetrabutylammonium fluoride (TBAF) in THF. Drying over  $\text{Na}_2\text{SO}_4$  followed by evaporation of the solvent afforded a mixture (45 mg) which was separated by PLC in benzene-acetone (8:2, v/v) to give five bands: 1 ( $R_F$  0.8, 3 mg), 2 ( $R_F$  0.54, 6 mg), 3 ( $R_F$  0.34, 5 mg), 4 ( $R_F$  0.29, 9 mg) and 5 ( $R_F$  0.18, 5.5 mg). The <sup>1</sup>H NMR spectrum of band 1 was identical to that of an authentic specimen of 3,5-dimethoxyphenol **27**.

Acetylation of band 2 and PLC in benzene-acetone (9:1, v/v) afforded 3',4',5,7-tetra-*O*-methyl-3-*O*-acetyl-ent-catechin **14** as a *white amorphous solid* ( $R_F$  0.63, 5 mg) (Found:  $M^+$ , 388.1521. Calculated for  $\text{C}_{21}\text{H}_{24}\text{O}_7$   $M$ , 388.1522;  $\delta_{\text{H}}$  (Table 2); CD  $[\theta]_{306.4}$  26,  $[\theta]_{279.4}$  4500,  $[\theta]_{267.6}$  1300,  $[\theta]_{251.6}$  -7.0,  $[\theta]_{243.9}$  -2400,  $[\theta]_{239.2}$  2.0,  $[\theta]_{232}$  5900,  $[\theta]_{221.1}$  790,  $[\theta]_{215.8}$  4400 and  $[\theta]_{210.4}$  -5.2.

Acetylation of band 3 followed by PLC in benzene-acetone (9:1, v/v) gave two bands, 3.1 ( $R_F$  0.55, 1.5 mg) and 3.2 ( $R_F$  0.43, 2.2 mg). Band 3.1 afforded 3',4',5-tri-*O*-methyl-3,7-di-*O*-acetylcatechin **16** as a *white*

*amorphous solid* (Found:  $M^+$ , 416.1463.  $C_{22}H_{24}O_8$  requires  $M$ , 416.1471);  $\delta_H$  (Table 2); CD  $[\theta]_{310}$  -300,  $[\theta]_{284}$  -340,  $[\theta]_{270.6}$  -1900,  $[\theta]_{255.6}$  -950,  $[\theta]_{235.6}$  -1300,  $[\theta]_{219.9}$  -46 and  $[\theta]_{210}$  3300. Band 3.2 comprised of the di-*O*-acetyl derivative **6** of the starting material **4**.

Band 4 was acetylated and the mixture was resolved by PLC in benzene-acetone (9:1, v/v) to give two fractions, 4.1 ( $R_F$  0.3, 2 mg) and 4.2 ( $R_F$  0.22, 5.4 mg). Fraction 4.1 afforded 3-*O*-acetyl-3',4',5,7-tetra-*O*-methylepicatechin-(2 $\beta$ ,1:4 $\beta$ ,2)-3-*O*-acetyl-4-[2*S*-acetoxy-3-(3,4-dimethoxyphenyl)propyl]-5-*O*-methylphloroglucinol **24** as a *white amorphous solid* (Found:  $M^+$ , 802.2839.  $C_{43}H_{46}O_{15}$  requires  $M$ , 802.2837);  $\delta_H$  (Table 1); CD  $[\theta]_{300}$  -290,  $[\theta]_{283.9}$  1000,  $[\theta]_{278.9}$  -53,  $[\theta]_{269.9}$  -23000,  $[\theta]_{259.5}$  -14000,  $[\theta]_{245.1}$  -66000,  $[\theta]_{232.5}$  -130,  $[\theta]_{225.6}$  33000,  $[\theta]_{217.2}$  60,  $[\theta]_{213.8}$  -15000,  $[\theta]_{210.5}$  -250,  $[\theta]_{207.7}$  9100,  $[\theta]_{205.1}$  7400 and  $[\theta]_{200.1}$  29000. Fraction 4.2 gave 2*R*,3*R*,4*R*,6*R*,7*R*-3,7-diacetoxy-9-methoxy-2,6-bis(3,4-dimethoxyphenyl)-4-(2-acetoxy-4,6-dimethoxyphenyl)-2,3-*cis*-3,4-*cis*-6,7-*cis*-3,4,7,8-tetrahydro-2*H*,6*H*-pyrano[2,3-*f*]chromene **8** as a *white amorphous solid* (Found:  $M^+$ , 802.2831.  $C_{43}H_{46}O_{15}$  requires  $M$ , 802.2837);  $\delta_H$  (Table 1); CD  $[\theta]_{300}$  -750,  $[\theta]_{280.7}$  8800,  $[\theta]_{262.2}$  370,  $[\theta]_{255.1}$  -580,  $[\theta]_{239.5}$  -75000,  $[\theta]_{221.6}$  120,  $[\theta]_{216.4}$  25000,  $[\theta]_{209.3}$  3100 and  $[\theta]_{202.6}$  20000.

Band 5 was acetylated and purified by PLC in benzene-acetone (9:1, v/v) to give 2*S*,3*R*,6*R*,7*R*-3,7-diacetoxy-9-methoxy-2,6-bis(3,4-dimethoxyphenyl)-2,3-*trans*-6,7-*cis*-3,4,7,8-tetrahydro-2*H*,6*H*-pyrano[2,3-*f*]chromene **20** ( $R_F$  0.4, 5 mg) as a *white amorphous solid* (Found:  $M^+$ , 608.2252.  $C_{33}H_{36}O_{11}$  requires  $M$ , 608.2258);  $\delta_H$  (Table 1); CD  $[\theta]_{310}$  -650,  $[\theta]_{256.6}$  -38,  $[\theta]_{234.6}$  -11000,  $[\theta]_{226.5}$  -9500,  $[\theta]_{212.4}$  -33000,  $[\theta]_{207}$  -1200 and  $[\theta]_{200}$  44000.

*Reductive cleavage of procyanidin A-1 hepta-O-methylether 3*. — The title compound (50 mg) was reduced with Na(CN)BH<sub>3</sub> (37 mg) and the resulting mixture worked-up as was described above for the procyanidin A-2 derivative **4**. This mixture (47 mg) was resolved by PLC in benzene-acetone (8:2, v/v) to give five bands: 1 ( $R_F$  0.8, 3 mg), 2 ( $R_F$  0.54, 7 mg), 3 ( $R_F$  0.33, 10 mg), 4 ( $R_F$  0.29, 20 mg) and 5 ( $R_F$  0.18, 33.5 mg). Band 1 again comprised 3,5-dimethoxyphenol **27**.

Acetylation of fraction 2 followed by PLC in benzene-acetone (9:1, v/v) afforded 3',4',5,7-tetra-*O*-methyl-3-*O*-acetyl-*ent*-catechin **14** ( $R_F$  0.63, 5 mg) with <sup>1</sup>H NMR (Table 2) and CD data identical to those of the same compound described above.

Fraction 3 was acetylated and the mixture resolved by PLC in benzene-acetone (9:1, v/v) to give two bands, 3.1 ( $R_F$  0.55, 2.5 mg) and 3.2 ( $R_F$  0.43, 2.2 mg). Band 3.1 gave 3,7-di-*O*-acetyl-3',4',5-tri-*O*-methylcatechin **18** as a *white amorphous solid* (Found:  $M^+$ , 416.1461.  $C_{22}H_{24}O_8$  requires  $M$ , 416.1471);  $\delta_H$  (Table 2); CD  $[\theta]_{310}$  -33,  $[\theta]_{290.4}$  -240,  $[\theta]_{281.8}$  790,  $[\theta]_{268}$  -1600,  $[\theta]_{256}$  -780,  $[\theta]_{242.9}$  -1800,  $[\theta]_{236.7}$  17,  $[\theta]_{231.6}$  2200,  $[\theta]_{225.4}$  -140,  $[\theta]_{220.4}$  1500,  $[\theta]_{214.4}$  70,  $[\theta]_{212.4}$  130 and  $[\theta]_{205.4}$  -1800. Band 3.2 afforded the di-*O*-acetyl derivative **5** of the starting material **3**.

Acetylation of fraction 4 followed by PLC in benzene-acetone (9:1, v/v) gave two bands, 4.1 ( $R_F$  0.32, 2 mg) and 4.2 ( $R_F$  0.21, 7 mg). Fraction 4.1 afforded the 2*R*-epimer **26** of compound **24** as a *white amorphous solid* (Found:  $M^+$ , 802.2823.  $C_{43}H_{46}O_{15}$  requires  $M$ , 802.2837);  $\delta_H$  (Table 1); CD  $[\theta]_{300}$  -410,  $[\theta]_{271.1}$  -14000,  $[\theta]_{260.5}$  -9900,  $[\theta]_{235.5}$  -81000,  $[\theta]_{228.5}$  -76000,  $[\theta]_{218.6}$  -470,  $[\theta]_{213.9}$  33000 and  $[\theta]_{206.4}$  -940. Fraction 4.2 gave the 7*S*-epimer **10** of the tetrahydropyrano[2,3-*f*]chromene derivative **8** as a *white amorphous solid* (Found:  $M^+$ , 802.2828.  $C_{43}H_{46}O_{15}$  requires  $M$ , 802.2837);  $\delta_H$  (Table 1); CD  $[\theta]_{295.8}$  -180,  $[\theta]_{279.8}$  13000,  $[\theta]_{261.5}$  420,  $[\theta]_{254.5}$  -1200,  $[\theta]_{242.4}$  -110000,  $[\theta]_{235.8}$  170,  $[\theta]_{233.5}$  12000,  $[\theta]_{229.4}$  9.5,  $[\theta]_{226.5}$  -5000,  $[\theta]_{223.7}$  -81,  $[\theta]_{217.3}$  18000,  $[\theta]_{213.5}$  190,  $[\theta]_{210.2}$  -190 and  $[\theta]_{207.6}$  -1800.

Acetylation of fraction 5 and subsequent PLC in benzene-acetone (9:1, v/v) afforded the 7*S*-epimer **22** of the tetrahydropyrano[2,3-*f*]chromene derivative **20** ( $R_F$  0.4, 5 mg) as a *white amorphous solid* (Found:  $M^+$ , 608.2251.  $C_{33}H_{36}O_{11}$  requires  $M$ , 608.2258);  $\delta_H$  (Table 1); CD  $[\theta]_{300}$  -210,  $[\theta]_{293}$  -500,  $[\theta]_{285}$  -2000,  $[\theta]_{281.1}$  -2000,  $[\theta]_{275}$  -2900,  $[\theta]_{258}$  -1600,  $[\theta]_{245}$  -11000,  $[\theta]_{238}$  -3.9,  $[\theta]_{231}$  1700,  $[\theta]_{227.4}$  1700,  $[\theta]_{213.9}$  5700 and  $[\theta]_{207.9}$  3900.

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### REFERENCES AND NOTES

1. Jacques, D.; Haslam, E.; Bedford, G.R.; Greatbanks, D. *J. Chem. Soc., Perkin Trans. 1*, **1974**, 2663.
2. Cronjé, A.; Burger, J.F.W.; Brandt, E.V.; Kolodziej, H.; Ferreira, D. *Tetrahedron Lett.*, **1991**, *30*, 3789.
3. Kolodziej, H.; Sakar, M.J.; Burger, J.F.W.; Engelshowe, R.; Ferreira, D. *Phytochemistry*, **1991**, *30*, 2041.
4. Gonzales, A.G.; Irizar, A.C.; Ravela, A.G.; Fernandez, M.F. *Phytochemistry*, **1992**, *31*, 1432.
5. Balde, A.M.; De Bruyne, T.; Pieters, L.; Kolodziej, H.; Van den Berghe, D.; Claeys, M.; Vlietinck, A. *Phytochemistry*, **1995**, *38*, 719; **1995**, *40*, 933, and references cited therein.
6. Karchesy, J.J.; Hemingway, R.W. *J. Agric. Food Chem.*, **1986**, *34*, 966.
7. Lane, C.E. *Synthesis*, **1975**, 135, and references cited therein.
8. Steynberg, J.P.; Burger, J.F.W.; Young, D.A.; Brandt, E.V.; Steenkamp, J.A.; Ferreira, D. *J. Chem. Soc., Perkin Trans. 1*, **1988**, 3323, 3331.
9. Steynberg, J.P.; Burger, J.F.W.; Young, D.A.; Brandt, E.V.; Ferreira, D. *Heterocycles*, **1989**, *28*, 923.
10. Steynberg, J.P.; Bezuidenhout, B.C.B.; Burger, J.F.W.; Young, D.A.; Ferreira, D. *J. Chem. Soc., Perkin Trans. 1*, **1990**, 203.
11. Inter-nuclear double resonance (INDOR) effects were avoided *via* multiple irradiation points for each on-resonance site - Kinns, M.; Sanders, J.K.M. *J. Magn. Res.*, **1984**, *56*, 518.
12. An NOE association between 10-H(D) and 2-H(C) in the  $^1H$  NMR spectrum of derivative **22** in  $C_6D_6$  permitted differentiation of both the aromatic and aliphatic spin systems and hence full assignment of the spectrum.
13. It should be emphasized that the stereochemistry of the flavan-3-ol derivative **13** from the reduction of the procyanidin A-1 derivative **3** is not compatible with the CD data of **3**. Such a discrepancy will be discussed in a forthcoming paper.
14. Beart, J.E.; Lilley, T.H.; Haslam, E. *J. Chem. Soc., Perkin Trans. 2*, **1985**, 1429.
15. Steynberg, P.J.; Steynberg, J.P.; Bezuidenhout, B.C.B.; Ferreira, D. *J. Chem. Soc., Chem. Commun.*, **1994**, 31; *J. Chem. Soc., Perkin Trans. 1*, **1995**, 3005.
16. Lewin, G.; Bert, M.; Dlauguet, J.-C.; Schaeffer, C.; Guinamant, J.-L.; Volland, J.-P. *Tetrahedron Lett.*, **1989**, *30*, 7049.
17. Burger, J.F.W.; Kolodziej, H.; Hemingway, R.W.; Steynberg, J.P.; Ferreira, D. *Tetrahedron*, **1990**, *46*, 5733.